was reported previously9. Investigation of the corpora allata ultrastructure with light microscopy (12-day-old bees treated on the 6th day and 25-day-old bees treated on the 15th day) revealed no degenerative changes after precocene treatment. From these experiments it can be concluded that precocene is not acting as anti-JH or as chemical allatectomizer in adult bees.

Egg-laying activity. The queens of 5 free flying colonies were treated topically with 30-200 µg precocene II. Weekly inspection of the brood area revealed no reduction of egglaying activity in comparison with the acetone controls during a period of 6 weeks after treatment. Also, compared with the controls, no size reduction was observed in the precocene treated colonies. Thus, it can be concluded that the egg viability was not affected by precocene application on the queen.

- The author thanks Dr A. Buehler for doing the histological part of the work, Dr H. Wille and Dr L. Gerig for valuable discussions and Dr S. Bogdanov for critical reading of the manuscript. The investigation was supported by the Swiss National Science Foundation grant No. 3.188.77.
- Roesch, G. A., Z. vergl. Physiol. 2 (1925) 571. Kratky, E., Z. wiss. Zool. 139 (1931) 120.
- 4 Nickel, K.H., and Armbruster, L., Arch. Bienenk. 18 (1937) 257.
- Lotmar, R., Landw. Jb. Schweiz 1939, 34.
- Lindauer, M., Z. vergl. Physiol. 34 (1952) 299.
- Maurizio, A., Landw. Jb. Schweiz 1954, 115. Merz, R., Gerig, L., Wille, H., and Leuthold, R., Revue suisse Zool. 86 (1979) 663.
- Fluri, P., Luescher, M., Wille, H., and Gerig, L., J. Insect Physiol. 28 (1982) 61.
- Rutz, W., Gerig, L., Wille, H., and Luescher, M., J. Insect Physiol. 22 (1976) 1485.
- Imboden, H., Wille, H., Gerig, L., and Luescher, M., Revue suisse Zool. 83 (1976) 928.
- Hardy, I., and Slama, K., Proc. 14th Int. Beekeep. Congr. 1963, p.49.
- van Laere, O., J. apic. Res. 13 (1974) 15.
- Engels, W., and Ramamurty, P.S., J. Insect Physiol. 22 (1976) 1427.

Mode of action of precocene II in the adult honey bee. Under natural conditions long life span worker bees have low JH titers9 while short life span is correlated with increased JH titers¹⁰. Thus the life shortening effect of high precocene doses cannot be interpreted as an anti-JH action. However, it cannot be decided from our experiments whether the life shortening effect of the high precocene dose is due to an anti-feeding action of the drug, followed by disturbance of endocrine processes²² or results from a general toxic effect. Furthermore it is not clear why queens can survive at much higher doses of precocene II than workers. Our results, together with those of other authors²³, might mean that the honey bees' corpora allata lack the enzymic capability for oxidizing precocene molecules to highly reactive epoxides which are supposed to exert a cytotoxic action on these glands.

- Fluri, P., Sabatini, A.G., Vecchi, M.A., and Wille, H., J. apic. Res. 20 (1981) 221.
- Imboden, H., Luescher, M., Revue suisse Zool. 82 (1975) 694.
- Bowers, W.S., Ohta, T., Cleere, J.S., and Marsella, P.A., Science 193 (1976) 542.
- Pratt, G.E., and Bowers, W.S., Nature 265 (1977) 548.
- Pener, M.P., Orshan, L., and de Wilde, J., Nature 272 (1978)
- 20 Unnithan, G.C., Nair, K.K., and Syed, A., Experientia 36 (1980) 135.
- Goewie, E.A., Beetsma, J., and de Wilde, J., Mitt. dtsch. Ges. allg. angew. Ent. I (1978) 304.
- Rembold, H., Czoppelt, Ch., and Sharma, G.K., Z. Naturforsch. C 34 (1979) 1261.
- Pratt, G.E., Jennings, R.C., Hamnett, A.F., and Brooks, G.T., Nature 284 (1980) 320.
- Lothmar, R., Beih. schweiz. Bienenztg. 1 (1945) 443.
- Ewen, A.B., J. Morph. 111 (1962) 255.

0014-4754/83/080919-02\$1.50+0.20/0© Birkhäuser Verlag Basel, 1983

Hyperglycemic effect in the rabbit induced by $ACTH_{4-10}$

R. Drouhault, A. M. Courtes and B. Dufy¹

Laboratoire de Neurophysiologie, Université de Bordeaux II, INSERM U.53, Domaine de Carreire, Rue Camille Saint-Saëns, F-33077 Bordeaux cedex (France), June 21, 1982

Summary. In the rabbit, a single i.v. administration of ACTH₄₋₁₀ (130 µg/kg) induces hyperglycemia. As ACTH₄₋₁₀ also induces hypocalcemia and inhibits insulin secretion, we suggest that hypocalcemia, inhibition of insulin secretion, and hyperglycemia, are closely related.

Studies of the relationship between structure and activity of adrenocorticotropin (ACTH) have shown that this hormone possesses different biological properties according to its various target cells. Since they share the same receptor in the adrenal gland the sequence ACTH₁₋₂₄ (synacthen) has the same biological action as the whole molecule.

It is known that the peptidic sequences necessary for the binding and/or the biological action are not always the same. The N-terminal sequence 1-10 of ACTH seems to be essential for the biological action, but the 11-24 sequence is necessary for the binding of the hormone on the adrenal target cells²⁻⁵. As a consequence of this observation numerous authors have reported that the 11-24 sequence, which is biologically inactive, can present an antagonistic effect towards the whole molecule^{4,6,7}. According to Beloff-Chain et al.8 the 18-39 sequence of corticotropin like intermediate

peptide (CLIP) should stimulate insulin secretion. On the other hand earlier studies have shown that in the rabbit $ACTH_{4-10}$ and in some instances α , β MSH or related pituitary peptides induce: a) hyperemia of adipose tissue⁹, b) increased concentration of free fatty acids (FFA) in adipose tissue, blood plasma, liver and kidney⁹, c) lipemia^{10,11}, d) acute nonkenotic metabolic acidosis^{12,13}, e) hypocalcemia with hypophosphatemia in rabbit but not in

In addition to these effects we show, in this paper, that ACTH₄₋₁₀ induces hyperglycemia and hypocalcemia, and inhibits insulin secretion.

Material and methods. Our experiments were performed on rabbits and rats. 'Fauve de Bourgogne' rabbits weighing 3000-3500 g, and Sprague-Dawley rats weighing 250-300 g, were kept without food for 24 h before the experiment but had free access to drinking water. The rabbits were divided into 3 groups of 8 animals. The 1st group was injected with 0.2 ml phosphate buffer (pH 6.2) containing 130 .µg/kg ACTH₄₋₁₀ (Bachem, Réf: 8688) via the marginal vein of the ear.

The 2nd group of animals received phosphate buffer only (pH 6.2). The 3rd group comprised untreated animals.

Venous blood samples (3 ml) were obtained by puncturing the marginal vein of the 2nd ear every hour for 4 h.

As a control, blood samples were taken from animals of all groups by puncturing the ear vein before any experimental manipulation. The rats were divided into 2 groups of 16 animals. The 1st group received in the tail vein 17 $\mu g/100$ g of ACTH₄₋₁₀ in the same phosphate buffer (pH 6.2). The 2nd group was injected only with phosphate buffer (pH 6.2). Four other untreated rats were used as control animals at time 0.

Four animals at a time were killed every hour by decapitation and blood samples collected in dry heparinized test tubes (for blood glucose) and dry test tubes (for Ca and insulin determination). a) Glycemia was determined by a colorimetric and enzymatic method with GOD-PAP kit (Boehringer-Mannheim), b) calcemia by atomic absorption technique using Pye Unicam SP 9, c) insulinemia by radioimmunoassay with a kit provided by C.E.A. (Commissariat à l'Energie Atomique Sorin Biomedica S.P.A.).

Results. In the rabbits $ACTH_{4-10}$ produced a significant hyperglycemia within 3 h. The control animals also showed an increase in plasma glucose although to a lesser degree than in $ACTH_{4-10}$ treated animals (fig. 1). Glycemia increased approximately 120% within 3 h (from 64 to 142 mg/100 ml). In the control animals this increase was only 65%. There was a significant difference (p < 0.05) between treated and control animals but none between the control animals and those treated with phosphate buffer only. Therefore, in the rabbits, puncturing the ear vein induced an appreciable hyperglycemia by itself. The time course of this effect in control animals was similar to that observed in

the treated ones. This may be due to an endogeneous peptide which is probably released under the stress produced by the puncture. In rats, glycemia of control animals was not different from treated animals and no increase in blood glucose was observed. These differences observed between rats and rabbits are comparable to those observed in studies of the effect of $ACTH_{4-10}$ on blood calcium. Whereas $ACTH_{4-10}$ induced a clear hypocalcemia in rabbit (fig. 2), this heptapeptide has no hypocalcemic effect in the rat¹⁴.

Plasma levels of insulin were decreased in ACTH treated rabbits (p < 0.05) (fig. 1). Conversely, rats did not show any similar modification of insulin levels following ACTH₄₋₁₀ injections. We observed that in all animals insulinemia regularly fell during the 3 1st h, increased during the 4th h and then slowly returned to normal values in the treated animals. This effect was greater in treated animals than in the control group. The time course of the effect was similar in both groups and seemed to be related to the variations in glycemia (fig. 1). In parallel experiments we investigated the dose response curve of insulin levels in response to ACTH₄₋₁₀ administration. The threshold for insulinemia was obtained with the administration of approximately 100 μ g/kg of ACTH₄₋₁₀ (fig. 3).

A putative nonspecific effect of the peptide was eliminated by the administration of an oxidation product obtained by treating the heptapeptide by performic acid which oxidates the methionine residue (situated in position 4). No significant difference on glycemia was observed compared to the control group in animals treated with such a compound. *Discussion*. In rabbits administration of ACTH₄₋₁₀ has been reported to increase plasma FFA levels, fatty liver and kidney lipemia^{10,15}. These changes have been shown to be direct and/or indirect consequences of FFA mobilization. In addition it has been clearly demonstrated that hypocalcemia stimulates the release of FFA from adipose tissue^{11,15-18}.

In the present study we report that administration of

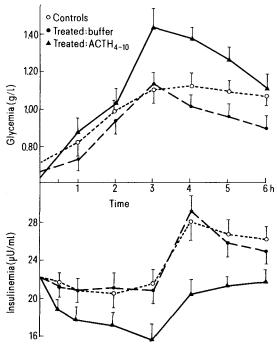


Figure 1. Effects of i.v. administrations of 130 μ g/kg of ACTH₄₋₁₀ on plasma glucose and circulating insulin levels. The determinations were made during 6 h. Mean values with SE determined in 6 animals.

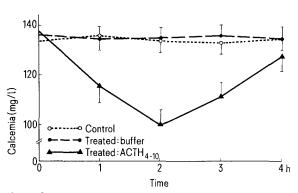


Figure 2. Effect of ACTH₄₋₁₀ (130 μ g/kg in 0.2 ml phosphate buffer) on circulating calcium levels in rabbits. Data are given as means \pm SE; n=6.

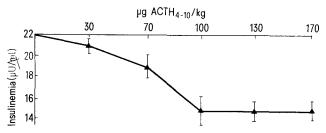


Figure 3. Dose response curve of insulinemia after i.v. administration of various amounts of ACTH₄₋₁₀. Data are given as means \pm SE; n = 6.

ACTH₄₋₁₀ to rabbits increased the plasma glucose concentration; this hyperglycemia was associated with a decline in insulin and calcium blood levels. In the rat, neither of these effects was observed. As ACTH is a hormone acting on the adrenal gland, the hyperglycemic effect observed after administration of $ACTH_{4-10}$ could be the consequence of a previous release of cortisol. This is not the case since ACTH₄₋₁₀, in contrast to ACTH, does not stimulate glucocorticoid production. Indeed, in our hands, injections of $ACTH_{4-10}$ up to 150 µg/kg were ineffective on blood cortisol levels. Our observations therefore agree with those of Smotherman et al.19 who obtained similar results in the rat. Moreover it has been shown by others and by us that this heptapeptidic sequence is inactive on the adrenal gland^{4,21,22}

Therefore we show that in the rabbit hypocalcemia, hypoinsulinemia and hyperglycemia are intimately related. In 1968, Curry et al.²³ showed the importance of calcium ions in insulin secretion. It can be suggested that hypocalcemia resulting from the administration of ACTH₄₋₁₀ in the

rabbit, or the release of an endogeneous peptide following stress in control animals, inhibits insulin secretion from pancreatic β cells, resulting in hyperglycemia. These results are in agreement with current findings on the role of Ca++ in the regulation of insulin release by pancreatic islets. The mechanism by which hypocalcemia could inhibit insulin release is beyond the scope of this short note. However it is clear that the insulin secretory process and the metabolism of glucose is controlled in β cells by calcium handling^{24–26}. Several mechanisms have been proposed; an action of the Ca⁺⁺ on enzymes converting proinsulin into insulin²⁶, an action on the microtubule filamentous system²⁷, or a modification of the cytoplasmic membrane permeability²⁶

The process by which ACTH₄₋₁₀ induces hypocalcemia in the rabbit is not yet clearly understood. However we suggest that ACTH₄₋₁₀ may stimulate the incorporation of calcium in various tissues resulting in a short term hypocalcemia. In agreement with this hypothesis we have demonstrated that ACTH₄₋₁₀ stimulates the uptake of Ca⁺⁺ in bone tissues²².

- We are grateful to Professor A. Baghdiantz for his helpful advice, critical reading of the manuscript and numerous helpful discussions during this study, and to Miss A. Ferret-Bouin for her technical assistance.
- Ramachandran, J., and Li, C.H., Adv. Enzym. 29 (1967) 391. Hofmann, K., Wingerder, W., and Finn, F.M., Proc. natl Acad. Sci. USA 67 (1970) 829.
- Saez, J.M., Dazord, A., Morera, A.M., and Bataille, P., J. Biol. Chem. 250 (1975) 1683.
- Seelig, S., Sayers, G., Schwyzer, R., and Schiller, P., FEBS Lett. 19 (1971) 232.
- Seelig, S., and Sayers, G., Archs Biochem. Biophys. 154 (1972)
- Finn, F.M., Widnell, C.C., and Hoffmann, K., J. biol. Chem. 247 (1972) 5695.
- Beloff-Chain, A., Edwarson, J.A., and Hawthorn, J., J. Endocr. 73 (1977) 28p.
- Rudman, D., Hirsch, R.L., Kendall, F.E., Seidman, F., and
- Brown, S.J., Rec. Prog. Horm. Res. 18 (1962) 89. Akgün, S., Rudman, D., and Wertheim, A.R., Endocrinology 84 (1969) 347.
- Tanaka, A., Pickering, B.T., and Li, C.H., Archs Biochem. Biophys. 99 (1962) 294.
- Rudman, D., and Shank, P.W., Endocrinology 79 (1966) 1100.
- Hoak, J.C., Poole, F.C.F., and Robinson, D.S., Circulation 28 (1963) 660 (Abstract).

- Drouhault, R., Valero, D., Baghdiantz, A., and Blanquet, P., J. Endocr. (1983) in press.
- Akgün, S., and Rudman, D., Endocrinology 84 (1969) 926.
- Anghileri, L.J., Experientia 15 (1975) 12. 16
- Schimmel, R.J., Biochem. biophys. Acta 326 (1973) 272. 17
- Yanagi, I., Okuda, H., and Fujii, S.S., J. Biochem. 66 (1969) 18
- 19 Smotherman, W.P., and Levine, S., Peptides 1 (1980) 207.
- Saez, J. M., Tell, G.E.P., and Riou, J.P., Lyon méd. 233 (1975) 20 1159.
- 21 Ramachandran, J., Farmer, S.W., Liles, S., and Li, C.H.,
- Biochim. biophys. Acta 428 (1976) 347. Drouhault, R., Valero, D., Basse-Cathalinat, B., Dupuy, J.C., 22 and Blanquet, P., Int. J. nucl. Med. Biol. 7 (1980) 281
- Curry, D.L., Bennett, L.L., and Grodsky, G.M., Am. J. Physiol, 214 (1968) 174,
- Malaisse, W.J., Hutton, J.C., Kawazu, S., Herchuelz, A., Walverde, I., and Sener, A., Diabetologia 16 (1979) 331
- Wollheim, C.B., Kikuchi, H., Renold, A.E., and Sharp, G.W.G., J. clin. Invest. 221 (1978) 451.
- Wollheim, C.B., and Sharp, G.W.G., Physiol. Rev. 61 (1981) 914.
- Howell, S. L., and Tyhurst, M., Diabetologia 22 (1982) 301.

0014-4754/83/080920-03\$1.50+0.20/0© Birkhäuser Verlag Basel, 1983

Different neuropathological effects of intrahippocampal injections of kainic acid and tetanus toxin

J. Kessler and H.J. Markowitsch^{1,2}

Department of Psychology, University of Konstanz, P.O. Box 5560, D-7750 Konstanz (Federal Republic of Germany), December 17, 1982

Summary. Behavioral and neuroanatomical effects of hippocampal injections of kainic acid (KA) and tetanus toxin (TT) were investigated in rats. Injections of KA resulted in both local and distant neuroanatomical damage, but not in clear signs of epilepsy; injections of TT on the other hand were followed (in some of the rats) by prolonged seizure attacks, but not by neuronal damage. Based on these results it is suggested that the widespread neuronal damage following KA lesions cannot be primarily attributed to orthodromic activation of epileptic discharges. Instead, specific properties of KA and their interactions with certain transmitters may provoke widespread neuroanatomical damage.

Neurotoxic agents are frequently used to study relations between brain damage and behavior³⁻⁵. Among these agents, kainic acid (KA) has been the most widely-applied substance in recent research. In spite of this fact, ideas on its mechanism in destroying nerve cells are still speculative and controversial⁶⁻¹³. The most common hypotheses on the action of KA in the brain are based on an 'excitoxic' action of KA^{7,10}: KA is thought to bind to specific receptors and thereby to induce an over-excitation of the neuron which ultimately may result in its destruction.

Because in experiments with KA a high correlation exists between the presence of epileptic discharges and the de-